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EARTH, OCEAN, AND ATMOSPHERIC SCIENCES

INVESTIGATING PARTICLE SIZE-FLUX RELATIONSHIPS AND THE BIOLOGICAL
PUMP ACROSS A RANGE OF PLANKTON ECOSYSTEM STATES FROM COASTAL TO
OLIGOTROPHIC

By
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TABLE OF CONTENTS

List of Figures	v
Abstract	vii
1. INTRODUCTION	1
2. METHODS	5
3. RESULTS	15
4. DISCUSSION	28
5. CONCLUSION	35
References	36
Biographical Sketch	47

LIST OF FIGURES

- Figure 1. Map of study region with bathymetry shown in gray scale and Lagrangian cycle tracks colored based on sediment trap measured export at a depth of 100 m ($\text{mg C m}^{-2} \text{d}^{-1}$).....6
- Figure 2. (A) Comparison of sediment trap and ^{238}U - ^{234}Th -based measurements of ^{234}Th flux for all Lagrangian cycles for cruises P0704, P0810, P1408 and P1604. Black line is 1:1 line. Non-parametric Spearman's $\rho = 0.51$ ($p = 4.9 \times 10^{-5}$). (B) Same as (A) but uses cruise averages for P1106 and P1208. Non-parametric Spearman's $\rho = 0.60$ ($p = 5.5 \times 10^{-5}$)16
- Figure 3. Comparison of flux estimates derived from sediment trap deployments and UVP algorithms using parameter values from (A) Guidi et al. (2008) and (B) Iversen et al. (2010) with shaded areas as the 95% confidence interval of an OLS regression determined via non-parametric bootstrapping. Each point is cycle averaged. Colors indicate vertically-integrated NPP. Grey dashed lines are 1:117
- Figure 4. (A) Size spectral slope of particles for the mixed layer (0-20 m) colored by sediment trap flux at 100 meters. (B) Same as (A) at each sediment trap deployment depth colored by the flux at that corresponding depth. Grey points are individual cast measurements, while colored points are cycle averages18
- Figure 5. Relative contribution of different particle sizes to flux if it is assumed that the Guidi et al. (2008) algorithm measures marine snow flux (red) and the experimentally measured recognizable fecal pellet flux (blue) is an additional flux term. (A) Cycle averaged UVP (Guidi) + fecal pellet flux estimates for CCEP0810 and CCEP1604, plotted against sediment trap flux. UVP (Guidi) + Fecal pellet flux as a function of particle size for (B) P0810-5 at 60 m, (C) P0810-6 at 100 meters, and (D) P1604-2 at 100 meters. (B, C, D) Subplot on the right shows the sum of UVP and fecal pellet flux (red and blue, respectively) compared to sediment trap flux (black).....20
- Figure 6. Representative images of particles collected in acrylamide gel sediment traps showing all particles (A, C, E), or magnified images of a subset of the particles (B, D, F). Images are from P1706-1, 150 m (A, B); P1706-3, 150 m (C,D); P1706-4, 50 m (E,F). (G) and (H) show representative images of the porous marine snow aggregates that were seen floating on top of the gels22
- Figure 7. Volume-weighted cumulative distribution function for particles that sank into acrylamide gel sediment traps on the P1706 cruise. Note that these do not include several large, porous marine snow aggregates that did not penetrate the gels23
- Figure 8. (A) Comparison of cycle averaged flux estimates derived from sediment trap deployments and the CCE-optimized UVP algorithm using fitted A and B values of 15.4 and 1.05 respectively with points colored based on primary productivity. The

calculated relative misfit (i.e. $(F_{UVP} - F_{ST}) / F_{UVP}$) plotted against (B) the Si:C ratio of sinking material, (C) the pigment:carbon ratio of sinking material, and (D) NPP.....25

Figure 9. (A) Blended ^{234}Th -sediment trap flux estimates from 50-200 meters compared to estimates predicted by the UVP algorithm using the optimized A and B values of 15.4 and 1.05 respectively. Points are colored by depth and the red dashed line is 1:1. (B) Misfit between the UVP flux estimate and the blended Th-234 method (i.e. $(F_{UVP} - F_{Th})$) plotted against depth, with a red line denoting the mean relative misfit (equivalent to AE) across all cruises26

Figure 10. Representative profiles of cycle averaged partial flux due to each particle size bin from 0m – 300m depth. Black dots represent Blended Th-ST flux estimates from 0-200 meters from cycles (A) P0810-6, (B) P1106-2, and (C) P1208-333

ABSTRACT

Sinking particles transport organic carbon produced in the surface ocean to the ocean interior, leading to net storage of atmospheric CO₂ in the deep ocean. The rapid growth of *in situ* imaging technology has the potential to revolutionize our understanding of particle flux attenuation in the ocean; however, estimating particle flux from particle size and abundance (measured directly by *in situ* cameras) is challenging. Sinking rates are dependent on several factors, including particle excess density and porosity, which vary based on particle origin and type. Additionally, particle characteristics are transformed while sinking. We compare optically-measured particle size spectra profiles (Underwater Vision Profiler 5, UVP) with contemporaneous measurements of particle flux made using sediment traps and ²³⁴Th:²³⁸U disequilibrium on six process cruises from the California Current Ecosystem (CCE) LTER Program. These measurements allow us to assess the efficacy of size-flux relationships for estimating fluxes from optical particle size measurements. We find that previously published parameterizations that estimate carbon flux from UVP profiles are a poor fit to direct flux measurements in the CCE. This discrepancy is found to result primarily from the important role of fecal pellets in particle flux. These pellets are primarily in a size range (i.e., 100 – 400 μm) that is not well-resolved as images by the UVP due to the resolution of the sensor. We develop new, CCE-optimized parameters for use in an algorithm estimating carbon flux from UVP data in the southern California Current ($\text{Flux} = \sum_{i=1}^x n_i A d_i^B \Delta d_i$), with A = 15.4, B = 1.05, d = particle diameter (mm) and Flux in units of mg C m⁻² d⁻¹. We caution, however, that increased accuracy in flux estimates derived from optical instruments will require devices with greater resolution, the ability to differentiate fecal pellets from low porosity marine snow aggregates, and improved sampling of rapidly sinking fecal pellets. We also find that the particle size-flux relationships may be different within the euphotic zone than in the shallow twilight zone and hypothesize that the changing nature of sinking particles with depth must be considered when investigating the remineralization length scale of sinking particles in the ocean.

CHAPTER 1

INTRODUCTION

Each year, approximately 40 - 50 Pg of carbon dioxide (CO₂) is fixed into organic matter in the ocean *via* photosynthesis (Le Quéré et al., 2018). The majority of this fixed carbon fuels the surface ecosystem and is quickly respired back into CO₂, which then equilibrates with the atmosphere. A small fraction of the organic matter produced by primary productivity escapes the euphotic zone and is transported to depth, primarily as sinking particles (Ducklow et al., 2001; Siegel et al., 2016). This process, known as the biological carbon pump (BCP), isolates carbon from the atmosphere for decades to centuries (Volk and Hoffert, 1985), and is estimated to transport between 5 and 13 Pg C from the euphotic zone each year (Henson et al., 2011; Laws et al., 2011; Siegel et al., 2014). Since marine photosynthesis accounts for about half of global photosynthesis (Field et al., 1998), the BCP is a key component in determining global and regional carbon budgets, which in turn are important for understanding climate change and for predicting environmental changes in future climate scenarios. Unfortunately, due to the numerous and complex processes that contribute to and influence the BCP, predicting its responses to climate change remains difficult (Passow and Carlson, 2012; Boyd, 2015; Burd et al., 2016).

The BCP is comprised of a suite of processes including active transport by vertically-migrating zooplankton and nekton (Steinberg et al., 2000; Hannides et al., 2009; Stukel et al., 2013; Davison et al., 2015; Kelly et al., 2019), subduction of refractory dissolved organic carbon (Carlson et al., 1994; Hansell et al., 1997), and subduction of particulate organic matter (Levy et al., 2013; Omand et al., 2015; Stukel and Ducklow, 2017; Llort et al., 2018), in addition to the flux of sinking particles (Martin et al., 1987; Buesseler and Boyd, 2009). However, sinking particles are typically assumed to dominate vertical flux relative to these other pathways of carbon transport (Turner, 2015; Stukel et al., 2018; Archibald et al., 2019). Quantifying the responses of the BCP to predicted changes in temperature, stratification, and surface wind stress thus requires sustained measurements of sinking particle flux across a wide range of marine ecosystems.

Sinking particle flux has traditionally been measured using sediment traps (Martin et al., 1987; Buesseler, 1991; McDonnell et al., 2015). These instruments have known issues associated with hydrodynamic biases, particle dissolution, and contamination by swimming zooplankton (Knauer et al., 1984; Baker et al., 1988; Lee et al., 1992; Buesseler et al., 2007). However, short-term deployments using surface-tethered traps with a large aspect ratio (height:diameter) and free-floating, neutrally-buoyant sediment traps seem to give accurate estimates of particle flux from the euphotic zone. Unfortunately, these short-term deployments require a substantial ship-time investment, because they typically require that a large research vessel remain in the vicinity of deployment for a period of days. This substantial cost has limited such time-series to only a few oceanic regions (Church et al., 2013; Lomas et al., 2013). Large, conical time-series traps, by contrast, offer an affordable approach to quantifying annual fluxes in the deep ocean (Honjo et al., 2008), but may exhibit extreme biases when deployed at shallow depths (Buesseler et al., 2010).

Clearly additional approaches are necessary in order to measure interannual variability in the global magnitude of the BCP. Radionuclide disequilibrium techniques (especially ^{238}U - ^{234}Th) provide one approach that requires substantially less ship-time investment (Buesseler et al., 1995; Van der Loeff et al., 2006; Waples et al., 2006). These methods exploit differences in activity between a conservative parent radionuclide and a particle-reactive, shorter-lived daughter isotope (Cochran et al., 2006; Waples et al., 2006). When the daughter particle is removed from the surface ocean through scavenging onto sinking particles a disequilibrium between parent and daughter is created. Measurement of this disequilibrium provides an estimate of radionuclide flux during a period of time related to the half-life of the daughter particle (Savoie et al., 2006). Radionuclide approaches are a powerful tool, because they measure a property (disequilibrium) that is directly created via particle flux and measurements can typically be made from a single CTD-Niskin rosette cast (Benitez-Nelson et al., 2001). However, substantial uncertainty can be introduced into carbon flux estimates from radionuclide disequilibrium due to high variability in carbon:radionuclide ratios over even relatively small distances (Buesseler et al., 2006; Hung and Gong, 2010; Stukel and Kelly, 2019). Furthermore,

radionuclide disequilibrium approaches are a moderately costly and time-intensive approach (at sea and post-cruise) and not suitable for autonomous measurements.

In recent years, the evolution of *in situ* optical/imaging approaches has opened up an indirect method for estimating particle flux (Jackson et al., 1997; Checkley et al., 2008; Picheral et al., 2010; Boss et al., 2015). These optical approaches permit high resolution sampling and are fully compatible with short station occupations. They enable particle flux estimation through either prolonged sampling durations that permit the observation of pulses of particles settling through the water column (Jackson et al., 2015), or, more commonly, through assumptions about the relationship between particle size and flux (Guidi et al., 2008; Iversen et al., 2010; Guidi et al., 2016; Ramondenc et al., 2016). However, there are substantial limitations associated with such approaches. Although theoretical arguments suggest a strong correlation between sinking speed and particle size, *in situ* measurements typically produce equivocal results (McDonnell and Buesseler, 2010; Jouandet et al., 2011). The poor correlation between sinking speed and size when measured *in situ* may result from the flux of <100- μm particles that are often not observed by optical instruments (Durkin et al., 2015) or as a result of the heterogeneity of sinking particles in the ocean and correspondingly different particle excess densities (Armstrong et al., 2009; Turner, 2015). Indeed, fecal pellets typically sink substantially faster than similarly sized marine snow aggregates (Stukel et al., 2014). Experimental results even show for a single particle class (abandoned appendicularian houses) that the aggregate size-sinking speed relationship changes with aggregate age (Lombard and Kiøboe, 2010; Lombard et al., 2013). There is thus good reason to believe that relationships between particle size and flux may be variable in time and space (Iversen et al., 2010; McDonnell and Buesseler, 2012).

In this study, we analyze results from six approximately month-long cruises in the California Current Ecosystem (CCE), during which we consistently deployed surface-tethered drifting sediment traps, quantified flux using ^{238}U - ^{234}Th disequilibrium, and measured profiles of particle size and abundance using the Underwater Vision Profiler 5 (UVP, Picheral et al., 2010). The CCE is an ideal study site because it offers a wide range of ecosystem conditions and plankton communities with commensurate impacts on the composition of sinking particles (Stukel et al., 2012; Ohman et al., 2013; Goericke and Ohman, 2015). By microscopically

quantifying fecal pellet flux and analyzing sinking particles and aggregates collected in polyacrylamide gels we further investigate the processes that alter the particle size-flux relationship. We then develop an algorithm, or more specifically a set of parameters to be used in an algorithm, optimized for the CCE that can potentially be used to estimate particle flux using both the UVP and fully autonomous instruments such as the *Zooglider* (Ohman et al., 2018). We present evidence that as our dataset encompasses a decade's worth of data in the CCE, including various frontal regimes, mesoscale, and submesoscale phenomena, these parameterizations are far more accurate to the region than values previously published. We caution, however, that more accurate quantification of flux from optical measurements likely requires additional information beyond particle size spectra, including characteristics related to the composition and identification of the particle or aggregate.

CHAPTER 2

METHODS

2.1 Data Collection

Data for this study come from seven process cruises for the CCE Long Term Ecological Research (LTER) program (P0704, April 2007; P0810, October 2008; P1106, June 2006; P1208, August 2012; P1408, August 2014; P1604, April 2016; P1706, June 2017; Figure 1). Each cruise was organized into quasi-Lagrangian experimental cycles (hereafter ‘cycles’) lasting 2-5 days (Figure 1). Cycle locations were chosen after an initial hydrographic survey of the epipelagic zone using a Moving Vessel Profiler (Ohman et al., 2012). Cycles were then initiated by the deployment of a surface-tethered drifting sediment trap (Stukel et al., 2013). This sediment trap and/or an independent array used for incubation of *in situ* experiments was then used as a moving frame of reference to enable prolonged study of the chosen water parcel (Landry et al., 2009). During the duration of the experiment, a suite of measurements was made including ^{14}C primary productivity (daily *in situ* incubations at 6 - 8 depths, Morrow et al., 2018), day-night paired net tows for mesozooplankton biomass and grazing (Landry et al., 2009), UVP measurements for zooplankton and particle vertical profiles (~10 casts per cycle, beginning with the P0810 cruise, Ohman et al., 2012; Biard et al., 2018), and ^{238}U - ^{234}Th disequilibrium (Stukel et al., 2019). A total of 33 different cycles were conducted as a part of this study, although 3 cycles did not include UVP data and 4 cycles do not have ^{234}Th data.

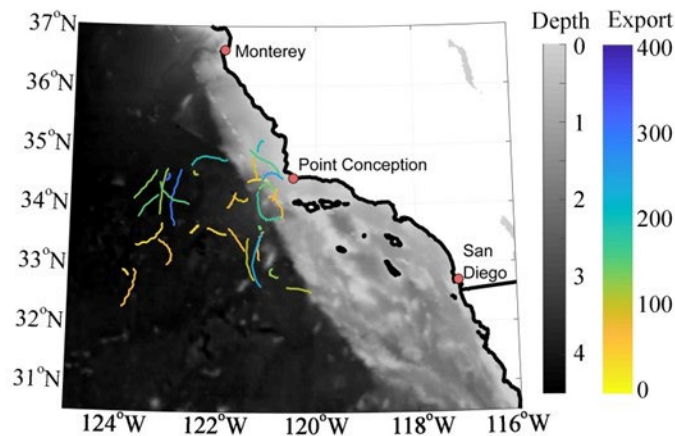


Figure 1. Map of study region with bathymetry shown in gray scale and Lagrangian cycle tracks colored based on sediment trap measured export at a depth of 100 m ($\text{mg C m}^{-2} \text{ d}^{-1}$).

Although we used identical methods on all cruises, the research foci of individual cruises varied leading to slightly different sampling strategies. During the P0704, P0810, P1408, and P1604 cruises our goal was to quantify ecosystem variability during El Niño neutral conditions (P0704 and P0810, Landry et al., 2012; Stukel et al., 2013) and during the anomalous 2014 warm conditions in the North Pacific and ensuing El Niño (P1408 and P1604, respectively; Kelly et al., 2018; Nickels and Ohman, 2018). On these cruises we thus sampled over a wide geographic region from water parcels near the Point Conception upwelling center to oligotrophic, offshore regions >400 km from shore. On the P1106 and P1208 cruises our focus was to investigate the impact of mesoscale fronts on ecological and biogeochemical dynamics (Krause et al., 2015; Stukel et al., 2017). Consequently, cycles were conducted over a more geographically restricted region, although ecosystem properties (phytoplankton biomass, primary productivity, mesozooplankton abundance, carbon export) varied substantially between the offshore side, coastal side, and core of these fronts. On the P1706 cruise our goal was to elucidate the role of mesoscale filaments in offshore transport of nutrients, carbon, and plankton communities. Cycles on this cruise were conducted over a reasonably large geographical area, as a result of the large spatial extent of the filament. However, these cycles can be considered a relatively coherent analysis of the temporal progression of a water parcel that is upwelled near the coast and ages over a period of weeks as it is advected from shore.

2.2 Sediment Traps

We used VERTEX-style drifting sediment traps with 69.85-mm inner diameter and an 8:1 aspect ratio (height:diameter) topped by a baffle constructed from smaller beveled tubes with a similar aspect ratio (Knauer et al., 1979). During P0704, cross frames consisting of 8 or 12 trap tubes were deployed at 100 m depth. On P0810, P1106, and P1208 cruises, traps were deployed at 100 m depth and also just below the base of the euphotic zone (if the euphotic zone was shallower than 75 m). On P1408, P1604, and P1706 cruises traps were deployed at the base of

the euphotic zone, 100 m, and 150 m. Trap tubes were filled with a hypersaline, poisoned brine solution (0.4% formaldehyde final concentration). Upon recovery, the overlying seawater was gently siphoned before the samples were gravity filtered through a 200- μm filter. This filter was then examined under a stereomicroscope to enable manual removal of swimming mesozooplankton taxa. After swimmer removal the <200- μm and >200- μm size fractions were recombined and samples were split for multiple analyses using a Folsom splitter. Samples for particulate organic carbon (POC) and nitrogen were filtered through pre-combusted glass fiber filters and frozen prior to fumigation with HCl and analysis with a CHN analyzer or isotope ratio mass spectrometer. Samples for C:²³⁴Th ratios were filtered through pre-combusted quartz (QMA) filters and analyzed as below. Chl-*a* and phaeopigments were measured by the acidification method (Strickland and Parsons, 1972). Samples for biogenic silica were filtered through a 0.6- μm polycarbonate filter and analyzed using a NaOH digestion procedure (Krause et al., 2015). A total of 33 different sediment trap deployments are included in this study and when individual depths are included, a total of 72 independent measurements of POC flux were made using sediment traps. See Stukel et al. (2013) and Morrow et al. (2018) for more details on sediment trap deployments and analyses.

On the P0704, P0810, and P1604 cruises, samples (typically ~1/2 tube) were stored in the trap brine (kept in the dark) and analyzed under stereomicroscope on land to enumerate and size mesozooplankton fecal pellets. Samples were placed in a settling chamber and fecal pellets were allowed to sink to the bottom for >6 hours. Water was then gently decanted through a 60- μm filter to ensure that none remained in the overlying water. Pellets were then transferred to a gridded petri dish and random grids were selected for analysis. Fecal pellets were identified by eye and imaged with a dedicated digital camera. From calibrated images, the length and width of each fecal pellet was recorded and a shape-specific volume and ESD were calculated. A shape-specific carbon to volume relationship (determined from CCE fecal pellet samples, Stukel et al., 2013) was applied in order to calculate the carbon content of each pellet.

On the P1706 cruise we also deployed sediment trap tubes containing a polyacrylamide gel designed to enable imaging of intact aggregates (Ebersbach and Trull, 2008; McDonnell and Buesseler, 2010). These trap tubes were identical to tubes used for other analyses, except that

they were unbaffled to ensure that there was no bias against >1-cm aggregates. The gel was contained in a custom-made acrylic chamber with an 82.55-mm inner diameter (i.e., slightly larger than the trap tube above it), because on previous cruises we found that material often clustered near the edge of the gel. Immediately after the cruise, a series of images covering the entire gel was taken under both bright field and dark field at 7.5X magnification (effective resolution 11.6 $\mu\text{m}/\text{pixel}$) with a Zeiss Discovery V20 stereomicroscope. 7.5X magnification bright field images were manually stitched together using Photoshop software. Particles were then manually circled and length, width, and area were extracted using Image J processing software (dark field images were simultaneously viewed to aid in particle and aggregate identification). A random set of locations on the gel was also imaged at 20X and 40X magnification. These higher magnification images were used only for qualitative analysis of smaller particles, because non-uniform loading of particles across the gel traps limited our ability to compute abundances from these images. Because the P1706 cruise had considerably higher particle flux than measured on previous cruises (beneath a coastal filament), particle loading at times obscured portions of the gel. Hence we were not able to image all of the particles on the gel and do not consider our analyses to give accurate estimates of the total flux of particles in any given size bin. Nevertheless, manual inspection of the images suggests no bias towards or against any particle size classes, so we believe that the slope of the particle size spectrum determined from the 7X magnification images of the acrylamide gels is accurate, although the intercept will not be.

2.3 $^{234}\text{Th}:$ ^{238}U Disequilibrium

Watercolumn ^{234}Th activity was measured using standard small volume methods (Pike et al., 2005). 4-L samples were drawn from Niskin bottles at typically 10-12 depths spanning the upper 200 m of the water column, with vertical resolution chosen to give higher resolution in the upper euphotic zone and/or near the deep chlorophyll maximum depending on water column structure. Samples were immediately acidified and spiked with a ^{230}Th tracer. After an equilibration period, NH_4OH was added to adjust to a pH of 8-9 and manganese chloride and

potassium permanganate were added. >8 hours later, manganese oxide precipitate was vacuum filtered onto a QMA filter, which was dried and mounted on a RISO sample holder. Beta activity was then measured with a RISO low background beta counter. After a final beta activity measurement >6 half-lives after collection, filters were digested in an 8M HNO₃ / 10% H₂O₂ solution, spiked with ²²⁹Th, and sonicated for 20 minutes. Th was then selectively isolated by ion chromatography, and isotope ratios (^{229:230}Th) were determined by inductively coupled plasma mass spectroscopy (at the Woods Hole Oceanographic Institution Analytical Lab or the National High Magnetic Field Laboratory) and used to determine ²³⁴Th yield. We determined ²³⁸U-²³⁴Th deficiency after estimating ²³⁸U activity from a linear salinity-uranium relationship (Owens et al., 2011). To quantify sediment trap sampling efficiency, we applied a one-dimensional steady-state equation without upwelling (Savoie et al., 2006). We then compared estimates of ²³⁴Th flux from the ²³⁸U-²³⁴Th disequilibrium approach to direct measurements of ²³⁴Th flux into sediment traps. C:²³⁴Th ratios determined from particles collected directly from sediment traps were then used in conjunction with the steady-state equation to derive carbon fluxes for the sediment trap depths.

2.4 Blended Flux Attenuation

Comparisons of sediment trap and ²³⁴Th results suggested no substantial over- or under-estimation of flux by either methodology (see results section 3.1.1). However, we note that the time-scale of flux measured by sediment traps (2-5 days) was matched with the time-scale of flux estimated via UVP (average of profiles determined over the same 2-5 days), while the time-scale of flux estimated by ²³⁴Th is approximately one month. Unlike sediment traps, the ²³⁴Th measurements can be used to determine continuous profiles of vertical flux equivalent to the vertical profiles of flux estimated by UVP. Consequently, we chose to create a blended export flux estimate that is equal to the sediment trap-derived flux at the deployment depths, while ²³⁴Th

measurements are used to extrapolate between sediment trap depths, above the shallowest trap deployment depth, and below the deepest deployment depth.

Assuming steady-state conditions without upwelling, carbon flux as a function of depth can be calculated from ^{234}Th - ^{238}U deficiency using the equation:

$$Flux(D) = CTh(D) \times \int_0^D \lambda_{234} \times Def(z) dz \quad (1)$$

Thus carbon flux attenuation (CFA) can be calculated as:

$$CFA = \frac{\partial(CTh(D))}{\partial z} \int_0^D (\lambda_{234} \times Def(z)) dz + \lambda_{234} \times Def(D) \times CTh(D) \quad (2)$$

where $CTh(D)$ is the $\text{C}:^{234}\text{Th}$ ratio of sinking particles at the depth horizon D , λ_{234} is the decay constant for ^{234}Th , and $Def(z)$ is the ^{238}U - ^{234}Th deficiency at depth z . Stukel et al. (2019) developed a robust algorithm for estimating the $\text{C}:^{234}\text{Th}$ ratio of sinking particles in the CCE from the ratio of vertically-integrated POC to vertically-integrated watercolumn ^{234}Th (${}^v\text{C}:^{234}\text{Th}_{tot}$):

$$\log_{10}(CTh(D)) = m \times \log_{10}({}^v\text{C}:^{234}\text{Th}_{tot}) + b \quad (3)$$

Combining Eqs. 1 and 2 allows us to quantify CFA and hence create smooth profiles of carbon flux above and below sediment trap depths. When sediment traps were deployed on two or more depths for the same cycle, we used $\text{C}:^{234}\text{Th}$ ratio measurements from those cycles to define the slope (m) in Eq. 3. When sediment traps were deployed at only a single depth, we used the CCE average slope calculated by Stukel et al. (2019): $m = 0.43 \pm 0.03$. In between sediment trap deployment depths, we stretched the CFA profiles (i.e., multiplied Eq. 2 by a constant) to ensure that the blended profiles exactly matched sediment trap-derived flux. Throughout the manuscript we are careful to differentiate between direct POC flux estimates made by sediment traps and blended POC flux estimates determined as above. For more details on calculating these blended profiles see Stukel et al. (2019).

2.5 UVP Measurements and Data Processing

The UVP was attached downward facing on the CTD Niskin-rosette, allowing for contemporaneous particle imaging and water column characterization, typically on 10 casts per cycle. For P0810, P1208, P1408, and P1604 the UVP5 standard definition model was used while for P1106 and P1706 a zooming version and HD model were deployed respectively. Each instrument was cross-calibrated in the lab prior to deployment. The UVP images a volume of ~1 L with each image and takes 6 images per second. It was typically deployed to a depth of 500 m. Post-cruise, data were processed using published Matlab scripts and uploaded to the Ecotaxa website (<https://ecotaxa.obs-vlfr.fr/>).

Particle data were obtained from the Particle Module of the Ecotaxa website. While the UVP reliably images organisms with a size >600- μm by converting the pixel border length to metric dimensions, it can also be used to quantify particle size and abundance down to the calibrated pixel size of the sensor, though for such small particles this is not as straight-forward a task. As a result of light-scattering, the relationship between the pixel area of particles and their corresponding metric area is nonlinear and follows a power law function (Picheral et al., 2010). This equation may be defined as $S_m = \alpha \cdot (S_p)^\beta$ where S_p is the surface of the particle in pixels, S_m is particle area in squared millimeters, and α and β are calibration constants for the UVP that were determined every year prior to deployment. Equivalent spherical diameter (ESD) was then calculated for each particle using this relationship. The unfiltered size range of particles measured by the UVP ranged from 42 μm to 81 mm, but to be consistent with previous literature we kept the upper size limit for flux calculations of 1.5 mm used by Guidi et al. (2008). As for the lower size limit, in addition to the limits imposed by camera resolution many of the exceptionally small signals were likely the result of noise or otherwise could not be reliably distinguished as actual particles, therefore raw particles were aggregated based on sampling year and used to plot particle size spectra (Supp. Figure 1). From these spectra the point at which the particle abundance relationship no longer followed a power law function was determined. However, because the camera and calibrations varied between cruises, there was no consistent size range at which the slope of the power law function changed, yet it was necessary to apply a uniform cutoff in order to determine a single parameter set representative of the entire dataset. 200 μm was chosen as the minimum particle size as it represented a reasonable compromise

when comparing the particle spectra and considering the importance of ~200 μm fecal pellets in sediment trap derived export (discussed further in Section 3.2). However, we note that, particularly for the 2008 cruise, the UVP is likely underestimating the abundance of 200-400 μm particles as the smallest particles detected were of 240 μm ESD. The remaining particles were then grouped into five-meter depth bins and one of 19 logarithmically spaced size bins based on their ESD. The volume sampled per image was multiplied by the number of unique values per depth bin (and therefore images) to get the total volume imaged, then divided by the range of the size class to obtain number of particles per liter.

Since both the mass and sinking speed of a marine snow aggregates can be related to the ESD of the aggregate by power law relationships (Alldredge and Gotschalk, 1988; Alldredge, 1998; Lombard and Kiørboe, 2010), Guidi et al. (2008) suggested that total particle mass flux (F [$\text{mg C m}^{-2} \text{d}^{-1}$]) can be calculated from the particle size distribution using the equation:

$$F = \sum_{i=1}^x n_i A d_i^B \Delta d_i \quad (4)$$

where n_i [$\text{L}^{-1} \text{mm}^{-1}$] is the particle number spectrum (i.e., the concentration of particles in bin i , divided by the bin width of bin i), d_i [mm] is the mid-point diameter of bin i , and A and B are constants related to the power law relationships that relate particle size to mass and settling velocity. Guidi et al. (2008) estimated values of $A = 12.5 \pm 3.4$ and $B = 3.81 \pm 0.70$ from 11 paired deployments of UVP and large conical sediment traps (typically two depths per deployment, with depths ranging from 100 to 1000 m, for a total of 23 independent flux measurements). Iversen et al. (2010) used a smaller number of deployments of similar large conical traps deployed at depths ranging from 1204 – 1866 m in the Mauritanian upwelling region and calculated different values for the constants in Eq. 2: $A = 273.8$ and $B = 4.27$. They suggested that these values may vary regionally.

We calculated flux from Eq. 4 based on the A and B values suggested by Guidi et al. (2008) and Iversen et al. (2010). In each instance, we averaged the flux estimates from the ~10 casts per Lagrangian cycle and compared this flux estimate to the sediment trap-derived POC flux measurements (a total of 69 independent sediment trap-derived flux POC were made on

cycles with associated UVP profiles). We also compared the UVP flux measurements to our blended flux profiles (separated into 5-m vertical bins from 50 to 200 m depth), which yielded 780 matchups between cycle average UVP flux and measured POC flux. In order to improve the model’s predictive capability for the CCE we calculated values of A and B specific to our dataset using a grid search method (to avoid the possibility of finding local, rather than global, minima). Following Guidi et al. (2008) and Iversen et al. (2010), we used a cost function equal to the square of the log-transformed differences between measurement and UVP-estimate:

$$\Delta F_C = \frac{1}{n} \sum_{i=1}^n \left(\log_{10}(F_{UVP,i}) - \log_{10}(F_{ST,i}) \right)^2 \quad (5)$$

where $F_{UVP,i}$ is the flux estimate for the i^{th} sediment trap measurement (note that this is an average of all the UVP measurements at that depth for a specific Lagrangian cycle), $F_{ST,i}$ is the flux measurement for sediment trap i , and n is the number of paired measurements ($n = 69$ for sediment trap-UVP comparisons). Although Guidi et al. (2008) and Iversen et al. (2010) used a lower threshold for particle size of 250 μm and 150 μm respectively (and we applied this limit when computing flux using their A and B values), we extended our particle size lower limit to 100 μm , when computing a CCE-optimized algorithm, because we found that small particles are particularly important to flux in the CCE.

2.6 Algorithm-data comparisons

In assessing the accuracy with which the UVP-dependent algorithms (i.e., models) approximate flux in the CCE, we treat sediment trap and sediment trap-blended *in situ* measurements as “truth”, but fully acknowledge that error also exists in these data (Lynch et al., 2009). Where possible, measurement uncertainty is included in figures as ± 1 standard deviation of the measurements, unless otherwise stated. Many different indices have been utilized to quantify model-data misfit (Stow et al., 2009). Following Guidi et al. (2008) and Iversen et al. (2010), we minimize the root mean log-transformed error (Eq. 5) when determining a CCE-optimized UVP flux algorithm. However, we also routinely report the average error

($AE = \frac{1}{n} \cdot \sum(model_i - data_i)$), the average absolute error ($AAE = \frac{1}{n} \cdot \sum(|model_i - data_i|)$), the average normalized absolute error ($ANAE = \frac{1}{n} \cdot \sum(|\frac{model_i - data_i}{model_i}|)$), and the Spearman's rank correlation (ρ). AE provides an estimate of the overall bias for a model. AAE gives an estimate of how far a model estimate typically is from reality, but is heavily dependent on uncertainty during high flux periods. The ANAE gives an estimate of the typical misfit divided by the model estimate and hence gives relatively equal weight to high and low flux conditions, while the MNAE gives an estimate of how far from the true value a typical model estimate is likely to be. Spearman's ρ quantifies how well the model and data covary.

CHAPTER 3

RESULTS

3.1 Comparison of Flux Estimates

3.1.1 Comparison of Sediment Trap and ^{238}U - ^{234}Th Deficiency Estimates

There was agreement between flux estimates made using sediment trap and ^{238}U - ^{234}Th deficiency approaches (Figure 2). When comparing ^{234}Th flux estimates made using ^{238}U - ^{234}Th deficiency and a one-dimensional steady-state approximation to ^{234}Th flux directly measured in sediment traps, we found strong agreement in the mean, with sediment trap measurements across all cycles and depths averaging 1% greater ^{234}Th flux than simultaneous ^{238}U - ^{234}Th measurements. The median of the ratio of sediment trap-derived flux to ^{238}U - ^{234}Th derived flux was 0.89. The sum of the squared log-transformed misfit between sediment trap and ^{238}U - ^{234}Th deficiency-derived flux was 0.086. (Note that since two values of ^{238}U - ^{234}Th deficiency-derived flux were negative and hence could not be log transformed, we replaced them with a value equal to 10% of the lowest positive ^{238}U - ^{234}Th deficiency-derived flux measurement). The AE was $-21 \text{ mg C m}^{-2} \text{ d}^{-1}$ and the ANAE was 0.19. The non-parametric Pearson's correlation coefficient (r) between sediment trap and ^{238}U - ^{234}Th deficiency-derived flux was 0.64 ($p < 0.001$). The weakest correlations between measurements occurred during the P1106 and P1208 cruises. This was not surprising, because on these cruises we focused on measurements within and immediately adjacent to strong mesoscale fronts. Residence time of water in these fronts (days) was substantially shorter than the temporal integration time-scale of ^{234}Th measurements (~ 1 month), but similar to the time-scale of sediment trap measurements (2 – 4 days). Cruise-average measurements determined by sediment traps and ^{234}Th were however, in close agreement for these two cruises (Figure 2B). When these cruise averages were used for P1106 and P1208 instead of the individual cycle averages, r increased to 0.76 ($p = 5.2 \times 10^{-8}$). This overall agreement gives us confidence that the sediment traps were accurately collecting sinking particles and that the mismatch between sediment traps and ^{238}U - ^{234}Th deficiency were primarily caused by differences in time scales and by occasional invalidation of the no-upwelling

assumption in the ^{238}U - ^{234}Th deficiency calculations. However, we cannot rule out the possibility of a sampling bias for any specific deployment.

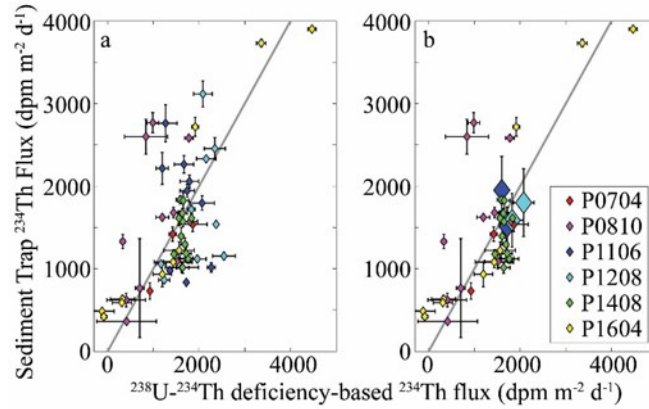


Figure 2. (A) Comparison of sediment trap and ^{238}U - ^{234}Th -based measurements of ^{234}Th flux for all Lagrangian cycles for cruises P0704, P0810, P1408 and P1604. Black line is 1:1 line. Non-parametric Spearman's $\rho = 0.51$ ($p = 4.9 \times 10^{-5}$). (B) Same as (A) but uses cruise averages for P1106 and P1208. Non-parametric Spearman's $\rho = 0.60$ ($p = 5.5 \times 10^{-5}$).

3.1.2 Comparison of Sediment Trap and UVP-Flux Estimates

In contrast to the sediment trap-thorium comparisons, we expect no mismatch in time scales between sediment trap and UVP-flux estimates. While the sediment traps integrate flux over the 2.25 to 5.25 day period of each Lagrangian experiment, the UVP estimates instantaneous flux during an individual profile. However, our UVP-derived flux measurements were based on approximately 10 casts per cycle, the first of which typically occurred less than one hour after sediment trap deployment and the last of which typically occurred ~ 4 hours prior to sediment trap recovery. Nevertheless, the sediment trap–UVP flux misfit was substantially greater than the sediment trap-thorium misfits, whether using the Guidi et al. (2008) or the Iversen et al. (2010) parameterizations (Figure 3). The Guidi et al. (2008) parameterization yields estimates substantially below the expected sediment trap flux, often by an order of magnitude. The AE for this algorithm was $-119.5 \text{ mg C m}^{-2} \text{ d}^{-1}$. This is a very large bias, considering that the average flux estimate from the algorithm was $42.1 \text{ mg C m}^{-2} \text{ d}^{-1}$. Indeed, the median flux estimated by this algorithm suggested a typical flux of $13 \text{ mg C m}^{-2} \text{ d}^{-1}$, which was lower than

the minimum sediment trap-measured carbon flux ($27 \text{ mg C m}^{-2} \text{ d}^{-1}$). This led to an ANAE equal to 6.0, suggesting that the true value was often 6-fold higher than that estimated by the Guidi et al. (2008) parameterization. In contrast, the Iversen et al. (2010) algorithm led to substantial overestimates of flux with $\text{AE} = 754.3 \text{ mg C m}^{-2} \text{ d}^{-1}$. Both the mean ($916 \text{ mg C m}^{-2} \text{ d}^{-1}$) and the median ($600 \text{ mg C m}^{-2} \text{ d}^{-1}$) flux estimates from the Iversen algorithm were higher than the carbon flux quantified during any of our sediment trap deployments.

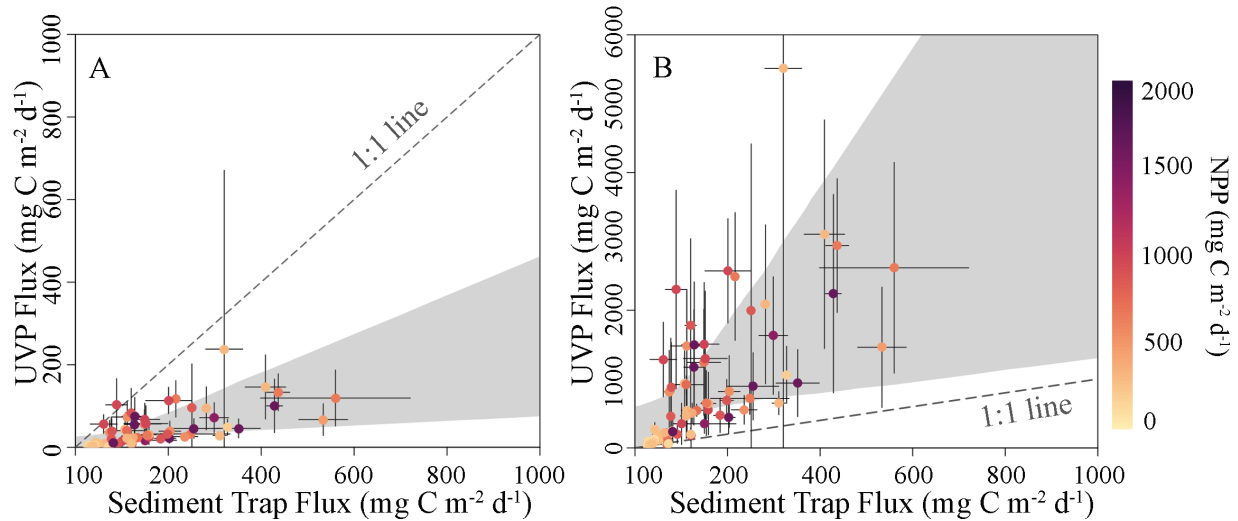


Figure 3. Comparison of flux estimates derived from sediment trap deployments and UVP algorithms using parameter values from (A) Guidi et al. (2008) and (B) Iversen et al. (2010) with shaded areas as the 95% confidence interval of an OLS regression determined via non-parametric bootstrapping. Each point is cycle averaged. Colors indicate vertically-integrated NPP. Grey dashed lines are 1:1.

3.2 Sinking Particle Size Spectrum

To investigate particle size variability and its relationship to flux, we computed the spectral slope of the modelled particle size spectrum for the mixed layer (to elucidate the relationship between mixed layer dynamics and sediment trap-derived flux) and for the specific depths of sediment trap deployments (to compare particle size at trap depth to sediment trap-derived flux directly). We found a slight though statistically-insignificant positive trend in the mixed layer (0-20 meters) between mean spectral slope and mean particle abundance ($\rho = 0.335$, $p = 0.081$; Figure 4A), potentially indicating that mean particle size increases when particle

abundance increases. This is likely the result of the dominance of large phytoplankton, particularly diatoms, during bloom conditions (Goericke, 2011b; a), which promote the presence of large zooplankton, large fecal pellets, and large aggregates that can all be detected by the UVP. Particle flux at 100 m depth was generally elevated when particle abundance and size spectral slope in the mixed layer were high. A weak potential trend was also seen when examining mean spectral slope as a function of mean particle abundance specifically for the depths at which sediment traps were deployed ($\rho = 0.242$, $p = 0.075$; Figure 4B). Interestingly, no relationship between mean spectral slope and sediment trap flux at corresponding depths was observed. However, particle flux was correlated with particle abundance at the same depth.

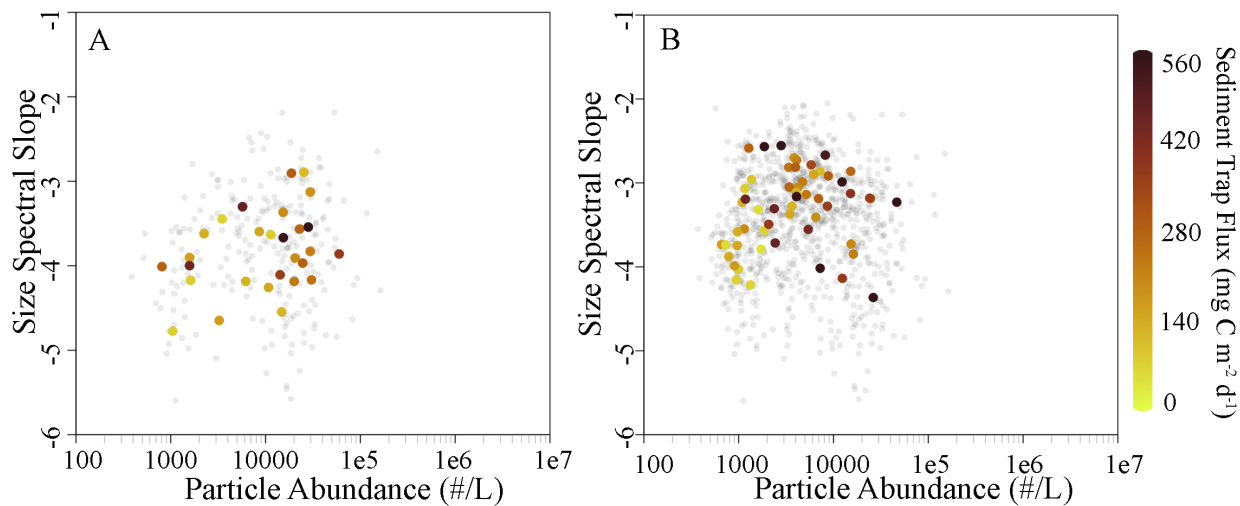


Figure 4. (A) Size spectral slope of particles for the mixed layer (0-20 m) colored by sediment trap flux at 100 meters. (B) Same as (A) at each sediment trap deployment depth colored by the flux at that corresponding depth. Grey points are individual cast measurements, while colored points are cycle averages.

To compare the particle flux size spectrum estimated by UVP to the sinking particles collected directly by sediment trap, we quantified the abundance and size of recognizable fecal pellets collected in sediment traps on the P0704, P0810, and P1604 cruises (Stukel et al., 2013; Stukel et al., 2018; Morrow et al., 2018). This estimate of recognizable fecal pellets is an underestimate of total fecal flux, because it excludes pellets that had been degraded such that

their original dimensions could not be determined, as well as pellets that were smaller than $\sim 60\text{-}\mu\text{m}$. However, we also note that it is possible that some of these pellets were transported to depth within larger aggregates that would register as large marine snow particles in the UVP. Across all cruises and cycles, 50% of fecal pellet carbon flux was mediated by fecal pellets $< 272\text{ }\mu\text{m}$ in ESD. This is notable for two reasons. First, fecal pellets often dominated flux into the sediment traps, particularly during high flux periods (Morrow et al., 2018). Second, the Guidi et al. (2008) and Iversen et al. (2010) algorithms explicitly exclude particles with an ESD less than $250\text{ }\mu\text{m}$ and $150\text{ }\mu\text{m}$, respectively, and find a relatively insignificant contribution to flux for all particles less than $500\text{ }\mu\text{m}$. This inherent mismatch is due to the assumption built into previous applications of the UVP-flux approach that large particles such as marine snow aggregates dominate carbon flux. Indeed, some studies have found that a substantial portion of carbon flux was dominated by $< 64\text{-}\mu\text{m}$ particles (Durkin et al. 2015; Hung et al. 2010). In our study, 32% of the fecal pellet fluxes was derived from pellets with an ESD $< 200\text{-}\mu\text{m}$, while 63% was derived from pellets with a width of $< 200\text{-}\mu\text{m}$. We find that if the flux of fecal pellets is added to the aggregate flux estimated by the Guidi et al. (2008) algorithm, we achieve a much better fit than if the Guidi et al. (2008) algorithm is used alone (Figure 5). While such an addition is surely an oversimplification and double-counts the $> 250\text{ }\mu\text{m}$ fecal pellets that are accounted for under the Guidi et al. (2008) algorithm, it illustrates the beginnings of a potential avenue for alleviating the apparent underestimation of flux. Furthermore, because of the large exponent found in the Guidi et al. (2008) algorithm, that algorithm assigns very little flux to particles in the size range of even large fecal pellets. There is, however, still a substantial misfit for cycles with low total flux. Despite the fact that recognizable fecal pellets were relatively few in these samples, the Guidi et al. (2008) algorithm substantially underestimated flux. These samples typically had a more negative size spectral slope (Figure 4). Hence it is possible that $< 60\text{ }\mu\text{m}$ fecal pellets were an important contributor to flux in these cycles or that smaller aggregates are more important than predicted by the Guidi et al. (2008) algorithm.

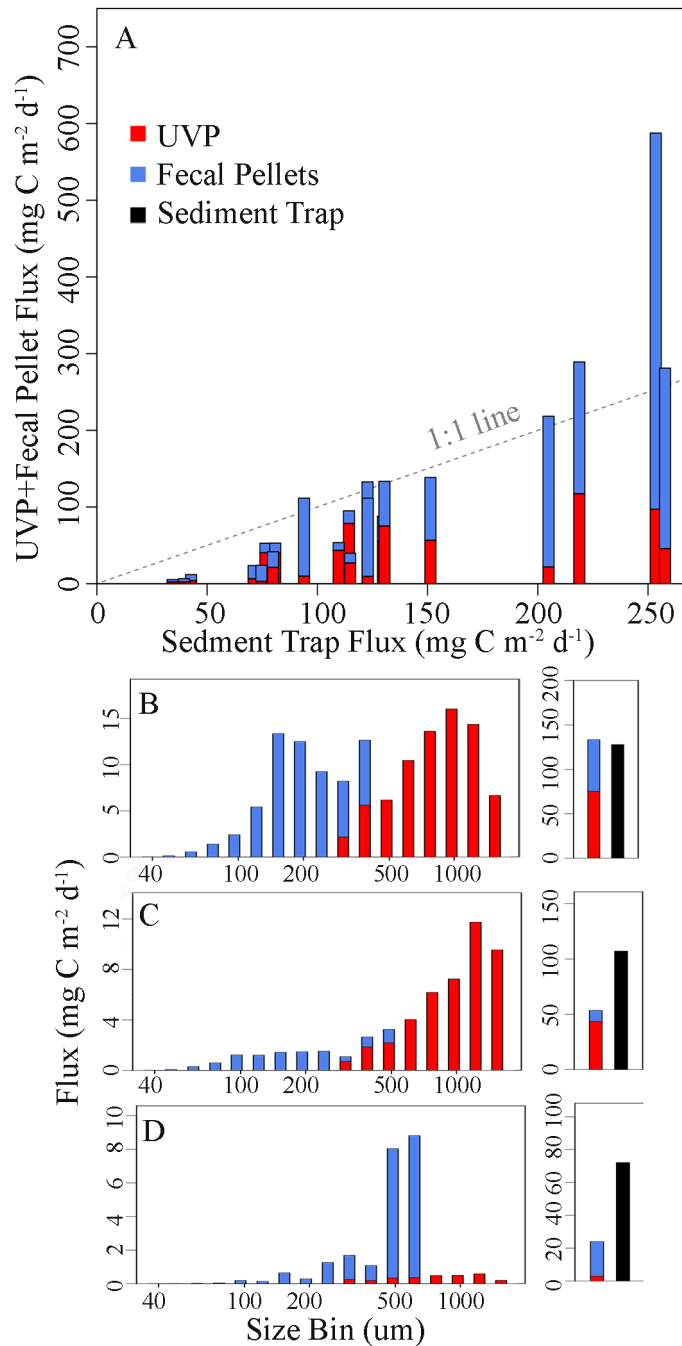


Figure 5. Relative contribution of different particle sizes to flux if it is assumed that the Guidi et al. (2008) algorithm measures marine snow flux (red) and the experimentally measured recognizable fecal pellet flux (blue) is an additional flux term. (A) Cycle averaged UVP (Guidi) + fecal pellet flux estimates for CCEP0810 and CCEP1604, plotted against sediment trap flux. UVP (Guidi) + Fecal pellet flux as a function of particle size for (B) P0810-5 at 60 m, (C) P0810-6 at 100 meters, and (D) P1604-2 at 100 meters. (B, C, D) Subplot on the right shows the

sum of UVP and fecal pellet flux (red and blue, respectively) compared to sediment trap flux (black).

To further investigate the size-flux relationship, on the P1706 cruise we utilized polyacrylamide gels located in the bottom of sediment trap tubes to collect particles and aggregates without destroying the structure of marine snow during sediment trap recovery (Figure 6). Materials seen in the gels were primarily of fecal pellet origin and had primarily sunk as independent pellets, rather than within marine snow particles. Total volume flux was typically dominated by particles in the $<600\ \mu\text{m}$ size range (Figure 7). In one sample (Cycle P1706-1, 150 m depth), $>50\%$ of volume flux was dominated by greater than 1 mm particles (Figure 7A, B); however, these large particles were not marine snow. They were identified as fish (likely anchovy) fecal pellets. In three samples (P1706-2, 50 m; P1706-3, 60 m; and P1706-4, 50 m) marine snow aggregates were also noted in the samples. However, they appeared to be less dense than the acrylamide gel and hence were not in the same focal plane as the rest of the particles. Therefore they were not included in the data assembled in Figure 6, but their impact will be further considered in the Discussion section. For P1706-2, three marine snow aggregates (equating to flux of $218\ \text{aggregates m}^{-2}\ \text{d}^{-1}$) were noted with an average ESD of 2.3 mm. For P1706-3, 24 marine snow aggregates ($1958\ \text{m}^{-2}\ \text{d}^{-1}$) were noted with ESD ranging from 1.6 – 7.1 mm. For P1706-4, an estimated 57 marine snow aggregates ($6550\ \text{m}^{-2}\ \text{d}^{-1}$) with typical ESD of 2.5 mm were noted. However, in this sample, the aggregates were clumped together making it difficult to get an accurate count. In all samples, these marine snow aggregates appear to be approximately spherical and highly porous (Figure 6G, H). Some contained phytoplankters and a few contained small fecal pellets. However, their contribution to total fecal pellet flux into the samples was negligible (likely $<1\%$), and we cannot rule out the possibility that the fecal pellets only became attached to the aggregates after sinking into the traps. Regardless, the high porosity of the aggregates, relative dearth of phytoplankton and fecal debris associated with them, and the high abundance of fecal pellets elsewhere in the samples, suggested that these marine snow particles were only minor contributors to the total carbon flux in all samples. No such marine snow aggregates were noticed in the acrylamide gel traps deployed at 150 m.

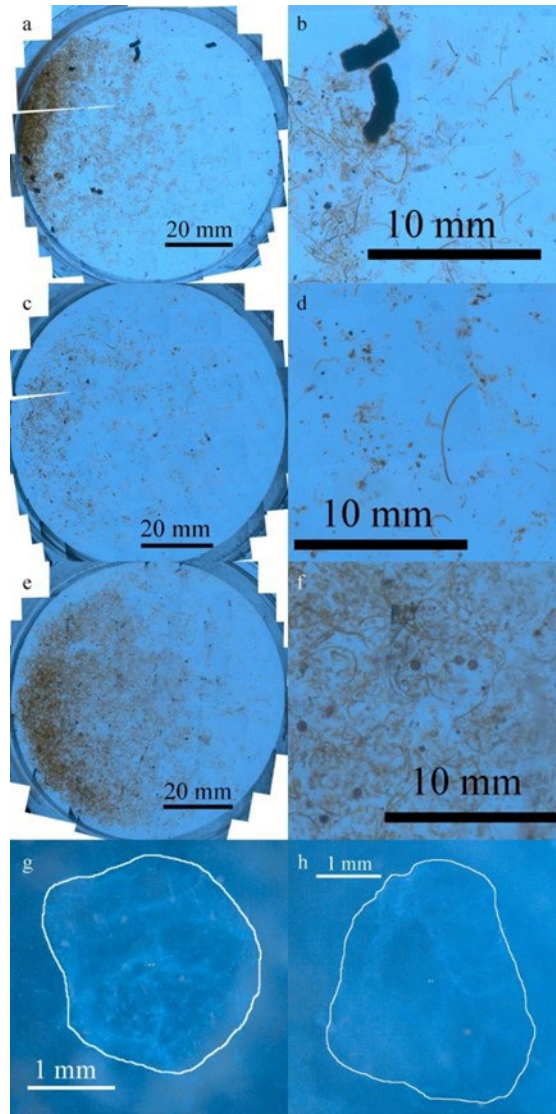


Figure 6. Representative images of particles collected in acrylamide gel sediment traps showing all particles (A, C, E), or magnified images of a subset of the particles (B, D, F). Images are from P1706-1,150 m (A, B); P1706-3, 150 m (C,D); P1706-4, 50 m (E,F). (G) and (H) show representative images of the porous marine snow aggregates that were seen floating on top of the gels.

Although there is a notable difference between the size relationships of volume-weighted flux estimates determined using acrylamide gels (mid-point of volume flux at $\sim 500 \mu\text{m}$) and the carbon-weighted flux estimates based on analyses of fecal pellets only (mid-point of mass flux at

~270 μm) we suspect that this discrepancy is derived from the different carbon:volume ratios of different classes of fecal pellets (large krill fecal pellets had lower carbon density than small copepod and appendicularian pellets), rather than any other difference resulting from the different methodologies. We caution, however, that all cycles of the P1706 cruise were associated with either the early, middle, or decline phases of a coastal diatom bloom and had above average carbon flux. Hence, these results do not inform the question of what the source or size of sinking particles during low flux periods may be.

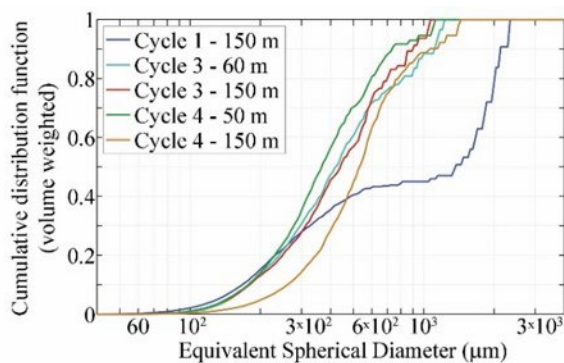


Figure 7. Volume-weighted cumulative distribution function for particles that sank into acrylamide gel sediment traps on the P1706 cruise. Note that these do not include several large, porous marine snow aggregates that did not penetrate the gels.

3.3 An optimized UVP algorithm for estimating flux in the CCE

3.3.1 Development of a CCE-optimized algorithm

We calculated CCE-optimized values for A and B in Eq. 4 that minimize the misfit to our sediment trap data as quantified in Eq. 5. Results suggest values of $A = 15.4$ and $B = 1.05$. Notably, our value for B is substantially lower than the values of 3.81 and 4.27 calculated by Guidi et al. (2008) and Iversen et al. (2010), respectively. B reflects the relative contributions of large and small particles to mass flux. Large values for B imply that large particles have a commensurately greater impact on mass flux relative to their standing stock as a result of increased mass, increased settling velocities, or a combination of the two. Since particle volume increases with ESD^3 , a value of $B < 3$ suggests that either the density or sinking speed of particles is inversely correlated with particle ESD.

The AE of our parameterization compared to sediment trap derived flux was $43.5 \text{ mg C m}^{-2} \text{ d}^{-1}$, with a median misfit of $-0.142 \text{ mg C m}^{-2} \text{ d}^{-1}$, suggesting that the algorithm had relatively low bias. The AAE and ANAE were $97.4 \text{ mg C m}^{-2} \text{ d}^{-1}$ and 0.6, respectively. Thus the CCE-optimized algorithm can be expected to under- or over-estimate flux by $97.4 \text{ mg C m}^{-2} \text{ d}^{-1}$ and the true value of flux should be expected to be within $\pm 60\%$ of the estimated value. These values suggest reasonably good model-data agreement, considering that the sediment trap estimates of carbon flux varied by a factor of 21 (from 27 to $560 \text{ mg C m}^{-2} \text{ d}^{-1}$), and that with the previously published algorithms, median export estimates were 77% lower (with the Guidi et al. 2008 algorithm) or 433% higher (with the Iversen et al. 2010 algorithm) than measured flux, while the CCE-optimized algorithm typically overestimated flux by only 16%.

3.3.2 Algorithm-data misfits

After the UVP carbon export estimates were obtained using the fitted parameters, the misfit (defined as UVP-ST flux) was examined against a variety of environmental parameters including primary productivity, depth, the slope of the size spectra, and the ratios of biogenic silicon, chlorophyll, phaeopigments, and nitrogen to organic carbon sinking material in order to evaluate covariance (Figure 8). In most cases these relationships were not significant. However, misfits were typically lower (i.e. UVP algorithm underestimated flux) when Si:C ratios were low ($\rho = 0.46$, $p = 0.004$; Figure 8B). This is the opposite of the effect we would expect if discrepancies were driven by increased silica-ballasting causing an increase in sinking speeds of similarly sized particles. This misfit also tended to be negative during cycles with the lowest primary production ($\rho = 0.312$, $p = 0.026$; Figure 8D).

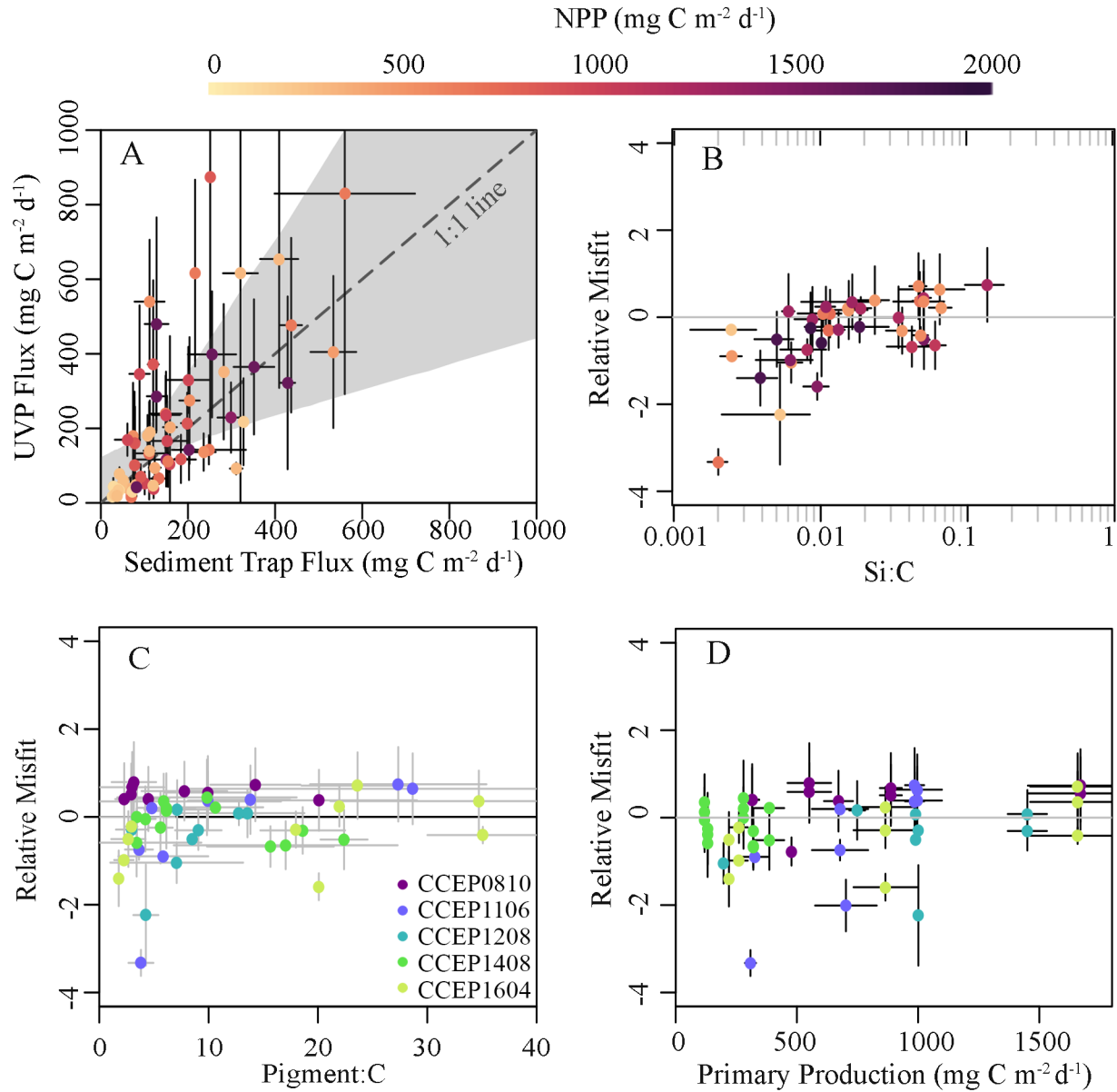


Figure 8. (A) Comparison of cycle averaged flux estimates derived from sediment trap deployments and the CCE-optimized UVP algorithm using fitted A and B values of 15.4 and 1.05 respectively with points colored based on primary productivity. The calculated relative misfit (i.e. $(F_{\text{UVP}} - F_{\text{ST}}) / F_{\text{UVP}}$) plotted against (B) the Si:C ratio of sinking material, (C) the pigment:carbon ratio of sinking material, and (D) NPP.

3.4 Particle flux vertical profiles

Comparison of our CCE-optimized UVP-derived flux estimates to flux profiles determined using a blended sediment trap- ^{234}Th approach (from 50 – 200 m depth) showed relatively good agreement (Figure 9A). There was a strong correlation between the two measurements (Spearman's $\rho = 0.59$, $p < 0.001$). The AE was $28.6 \text{ mg C m}^{-2} \text{ d}^{-1}$, showing no substantial bias in the data overall, while the ANAE was 0.68. However, over the 50 – 200 m depth range we did find a slight difference in algorithm accuracy with depth (Figure 9B). The AE was $112.5 \text{ mg C m}^{-2} \text{ d}^{-1}$ at 50 m, $39.9 \text{ mg C m}^{-2} \text{ d}^{-1}$ at 100 m, $12.5 \text{ mg C m}^{-2} \text{ d}^{-1}$ at 150 m, and $108.9 \text{ mg C m}^{-2} \text{ d}^{-1}$ at 200 m. This suggests that (at least over the limited depth range of the upper twilight zone) changing relationships between particle size and flux do not lead to major biases in profiles of vertical flux. However, manual inspection of profiles showed that for most cycles there was a depth near the base of the euphotic zone at which UVP-derived particle flux estimates substantially overestimated measured flux (Figure 10). Since UVP flux estimates were based on particle size and abundance, this suggests that near the base of the euphotic zone there is a large abundance of particles that are sinking more slowly than similarly-sized particles deeper in the euphotic zone.

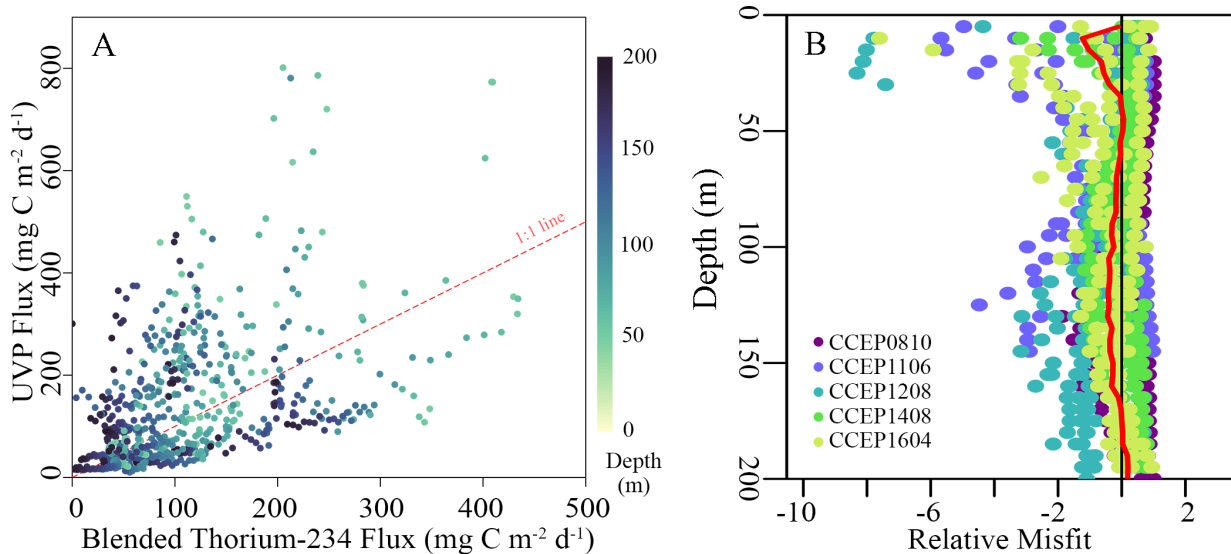


Figure 9. (A) Blended ^{234}Th -sediment trap flux estimates from 50-200 meters compared to estimates predicted by the UVP algorithm using the optimized A and B values of 15.4 and 1.05

respectively. Points are colored by depth and the red dashed line is 1:1. (B) Misfit between the UVP flux estimate and the blended Th-234 method (i.e. $F_{\text{UVP}} - F_{\text{Th}}$) plotted against depth, with a red line denoting the mean relative misfit (equivalent to AE) across all cruises.

CHAPTER 4

DISCUSSION

4.1 Quantifying vertical carbon flux

Many marine ecosystems are rapidly responding to a changing climate (Behrenfeld et al., 2006; Doney et al., 2012; Asch, 2015). Whether these changes will lead to increased or decreased efficiency of the BCP ultimately depends on changes in nutrient supply (Bakun, 1990; Garcia-Reyes and Largier, 2010; Rykaczewski and Dunne, 2010; Capotondi et al., 2012; Sydeman et al., 2014) and the ways in which plankton communities adapt to altered physical forcing (Richardson and Schoeman, 2004; Montes-Hugo et al., 2009; Boyd, 2015; Cael et al., 2017). The complex interactions of different components of the BCP make predictions of future changes in the BCP highly uncertain. As a result, sustained large-scale measurements of carbon flux are a research priority. Unfortunately, the most direct approach for measuring sinking particles (i.e., sediment traps) is not well suited to long-term deployments in the upper water column, where moored time-series traps may be unreliable (Buesseler et al., 2010), and surface-tethered or neutrally-buoyant traps require sustained research vessel presence.

The development of methods appropriate for use on survey or monitoring cruises and/or autonomous deployments is thus an important research focus (McDonnell et al., 2015). Our results suggest that ^{238}U - ^{234}Th deficiency measurements provide a more robust approach for estimating particle flux than UVP-based estimates. Despite not being calibrated to the CCE region, the thorium approach showed no significant bias with respect to estimation of particulate ^{234}Th fluxes (although we note that *in situ* pump values of the C: ^{234}Th ratio, as would normally be used to convert to carbon fluxes on a survey cruise, underestimated sinking particle C: ^{234}Th ratios in the CCE by an average of 44% (Stukel et al., 2019)). The greater accuracy of the ^{238}U - ^{234}Th deficiency approach is not surprising, since it estimates carbon flux from a direct *flux* measurement (i.e. ^{234}Th flux), while the UVP-based approach infers flux from measurements of particle *standing stock*. Nevertheless, the increased cost, required expertise, and added water budget requirements of ^{234}Th measurements suggest that they are not likely to be as widely used as optical particle measurements. Furthermore the suitability of optical sensor deployment for

autonomous deployments highlights the utility of algorithms that can quantify flux from such measurements, even if these results require careful (and potentially sustained) tuning and validation.

Most optical approaches rely on a strong covariance between particle size (or a proxy for particle size) and the property of interest, which for carbon export is mass flux. Previous work has demonstrated trends between particle size and sinking velocity (Smayda, 1971; Alldredge and Gotschalk, 1988; Stamieszkin et al., 2015), as well as particle size and mass (Alldredge, 1998). Therefore a reasonable approach was undertaken in Guidi et al. (2008) to couple these properties into mass flux for the UVP platform. However, the use of previously published parameterizations for estimating flux from UVP data led to substantial misfits with our measurement data. With the Guidi et al. (2008) global algorithm, the UVP flux estimate was biased low, often underestimating actual flux by $\sim 90\%$. The misfit was particularly stark during low-biomass, low-flux periods when the UVP often predicted flux $< 7 \text{ mg C m}^{-2} \text{ d}^{-1}$, even though sediment trap measured flux was never $< 26 \text{ mg C m}^{-2} \text{ d}^{-1}$. In contrast, the Iversen et al. (2010) parameterization led to substantial overestimates of flux often by a 10:1 ratio, with misfits that were greatest when particle loading was highest. This parameterization suggested that carbon flux should exceed $1000 \text{ mg C m}^{-2} \text{ d}^{-1}$ for 21 (out of a total of 69) paired sediment trap-UVP measurements. Measured sediment trap flux never exceeded $560 \text{ mg C m}^{-2} \text{ d}^{-1}$. These results echo the conclusions of Iversen et al. (2010) that UVP-flux estimates need to be tuned to specific regions. This suggests that extreme caution should be utilized when attempting to quantify flux from UVP-profiles using a global algorithm. Although Iversen et al. (2010) suggested that the Guidi et al. (2008) algorithm may work better in offshore regions than coastal environments, we actually find the greatest mismatch to occur during oligotrophic conditions.

When optimizing Eq. 4 to the CCE domain, we found a much lower exponent ($B = 1.05$) than that obtained by either Iversen et al. (2010) or Guidi et al. (2008). This B value is similar to the theoretical B value calculated by Iversen et al. (2010) from relationships between aggregate size-mass and size-settling velocity relationships published in Alldredge and Gotschalk (1988). Nevertheless, this exponent is surprisingly low for a system in which fecal pellets are a dominant contributor to flux and for which limited evidence suggests that carbon density decreases only

slightly with volume (Stukel et al., 2013). Indeed, given reasonable exponents for the scaling relationship between fecal pellet ESD and mass (>2), a value of $B = 1.05$, would imply that larger pellets sink more slowly than smaller pellets. This result would be in stark contrast with multiple theoretical and experimental studies suggesting that larger fecal pellets sink faster (Small et al., 1979; Turner, 2002; Giesecke et al., 2010). Instead, we suspect this small exponent may reflect the fact that flux was dominated by particles with sizes near the lower size detection limit of the UVP (i.e., 42 μm – 125 μm depending on calibration). If 200 μm to 300 μm particles are dominating flux, the UVP will struggle to accurately differentiate their sizes. This difficulty is exacerbated by the non-spherical shape of fecal pellets, which had typical length:width ratios ranging from 2:1 to 10:1 (Stukel et al., 2018). Thus the abundance of such particles (rather than slight differences in their sizes that may not be accurately recorded by the UVP) becomes a dominant predictor of flux. The fecal pellet data presented here illustrate an important caveat for imaging systems. Imaging systems are well suited to observing objects over a range of approximately 3 orders of magnitude (i.e. 10 pixels to $\sim 1,000$ pixels in length), yet particles within the water column often range over 5 orders of magnitude from <1 μm to >10 cm. The pixel size on the UVP5 thus results in coarse granularity of particles in regions where fecal pellets are abundant and important to mass flux. One potential solution to this that is increasingly being implemented in the field is the use of multiple instruments with varying resolutions to adequately cover the full size range of marine particles (e.g., Jackson et al., 1997). Such increased resolution would allow accurate particle shape (and potentially color) information to be used in classifying particles and more accurately determining volume and carbon content.

Our results thus suggest that, in a region dominated by fecal pellet flux, more accurate estimates of flux will require optical profiling instruments with finer resolution at small particle sizes. Furthermore, in regions where the relative contribution of different particle classes (e.g. phytodetritus, fecal pellets, mucous feeding webs, or marine snow of mixed origin) can vary substantially, optical approaches may need to incorporate information other than size (shape, color, porosity) into flux calculations. Alternately, the use of autonomous ‘optical sediment traps’ may provide a feasible approach for estimating flux driven by heterogeneous particle classes, because the ability of such approaches to actually quantify particle flux onto a glass plate

obviates the need for an assumed particle size-settling velocity relationship (Bishop et al., 2016; Estapa et al., 2013; Estapa et al., 2019). However, such approaches still rely on accurate determination of carbon density from particle optical properties (Estapa et al., 2017).

4.2 Vertical particle flux and attenuation

Our results shed light on the processes driving patterns in vertical POC flux in the CCE. Fecal pellets in the $\sim 100 \mu\text{m} - 400 \mu\text{m}$ size range appear to be the dominant contributors to export flux in the shallow twilight zone (Figures 5-7). Flux of these pellets shows substantial spatial variability, with greater fecal pellet (and total carbon) flux beneath coastal, bloom water parcels than beneath the oligotrophic waters typically found offshore. The prevalence of small particles in flux leads to a low exponent ($B = 1.05$) for the relationship between particle size and mass flux. This stands in marked contrast to exponents determined in other regions, such as the Mauritanian Upwelling region, where B values >3 (and *in situ* estimates of the particle size-spectrum) suggest a greater importance for mm-sized marine snow aggregates (Iversen et al., 2010). Notably, the Mauritanian Upwelling region is subject to substantial Saharan dust deposition, which may contribute to mineral ballasting and higher settling velocities (Iversen et al., 2010; van der Jagt et al., 2018).

Such large aggregates were in fact found in acrylamide gels placed in the bottom of sediment traps on our P1706 cruise. They would have contributed a substantial amount of the volume of sinking material in the samples if they had penetrated the gels sufficiently to enable their quantitative analysis. However, their density appeared to be less than that of the acrylamide gel, resulting in them essentially floating atop the gel. Despite their volume, however, visual inspection of this marine snow suggested that they had only a marginal contribution to carbon flux, as a result of high porosity and a paucity of visible material (fecal pellets and phytodetritus) within the aggregates. These marine snow particles were only seen in acrylamide gel traps deployed at a depth of 50 m – 60 m, however, and were absent from traps at 150 m. Taken together this suggests that: 1) these aggregates likely have only moderate sinking rates as a result of high porosity and correspondingly low excess density, 2) these aggregates are likely a

substantial contributor to *volume* flux near the base of the euphotic zone, but only minor contributors to *carbon* flux at this depth, and 3) these marine snow aggregates experience substantial flux attenuation in the shallow euphotic zone.

The results above provide an interesting interpretation of contrasting results determined using alternate approaches to quantify export in the CCE region. Jackson & Checkley (2011) used an autonomous laser optical plankton counter to quantify particle abundance, volume, and flux (assuming Stokes' Law relationships and constant carbon:volume ratios). They concluded that flux near the base of the euphotic zone was quite high (with maximum values near $800 \text{ mg C m}^{-2} \text{ d}^{-1}$), but that flux attenuation was exceedingly rapid in the shallow twilight zone, and decreased to $\sim 10\%$ (sometimes substantially less) by a depth of 100 m. From these results, the authors concluded that the base of the euphotic zone was a region where zooplankton play a key role as gatekeepers of flux into the mesopelagic. Our sediment trap and ^{234}Th measurements, however, show no such zone of high carbon flux attenuation (Figure 10). Rather, our combination of direct flux measurements and *in situ* particle imaging paint a more nuanced picture of particle transformations in this dynamic depth domain. We find that these depths are precisely the depths where our CCE-optimized algorithm consistently overestimates flux relative to sediment traps and ^{234}Th . This result strongly suggests that the relationship between size and sinking speed shifts near the base of the euphotic zone, with rapidly-sinking particles (within any given size range) contributing more to flux at deeper depths than slowly-sinking particles. Indeed, Guidi et al. (2008) utilized sediment trap deployments between 150 and 1000 m for their parameterizations as opposed to the present study which relied on data from 50-150 m, which may in part explain their drastically different values of B.

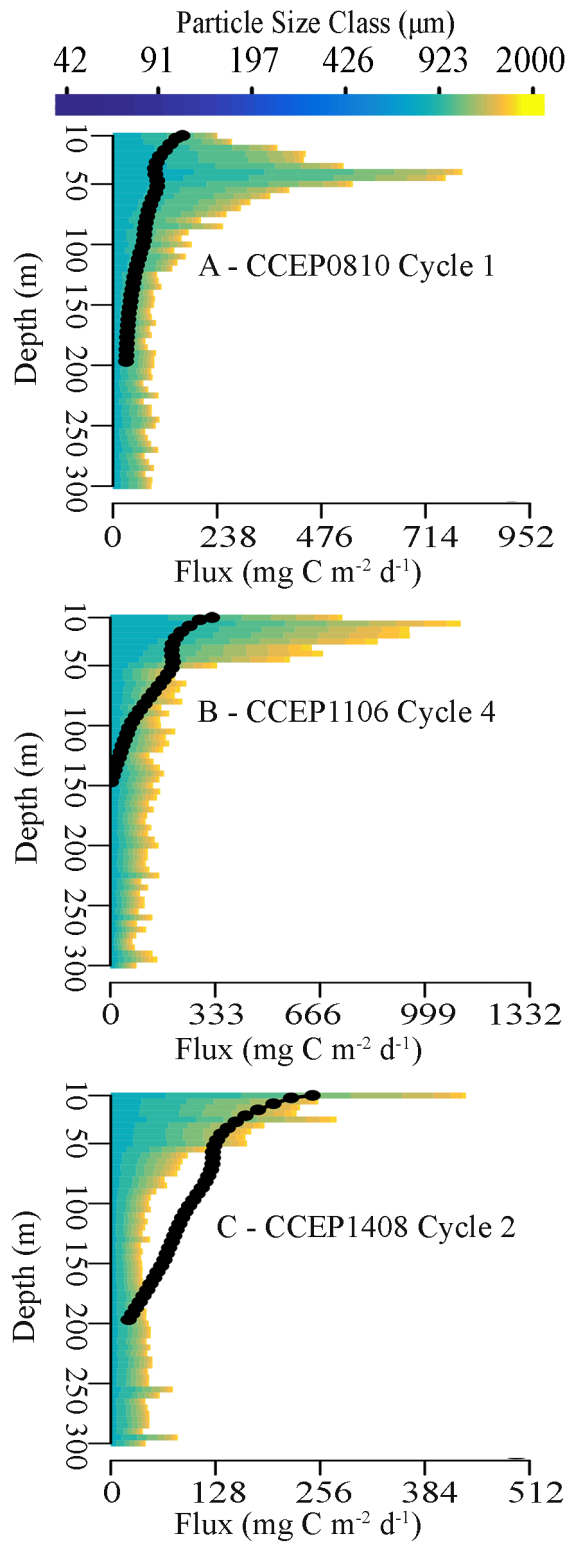


Figure 10. Representative profiles of cycle averaged partial flux due to each particle size bin

from 0m – 300m depth. Black dots represent Blended Th-ST flux estimates from 0-200 meters from cycles (A) P0810-6, (B) P1106-2, and (C) P1208-3.

There are many potential possibilities for explaining such discrepancies. As hypothesized by Jackson and Checkley (2011), zooplankton may play an important role at this depth, particularly if they are suspension-feeders with high clearance rates that remove most of the slowly sinking particles before they reach deeper depths (Stukel et al., 2019). Sinking rates of low excess density marine snow may also be retarded at density gradients found near the base of the mixed layer (MacIntyre et al., 1995; Prairie et al., 2015). Aggregate shape, size, and sinking speed can also change with age due to physical and biotic forces (Plough et al., 2008; Lombard and Kiørboe, 2010). Particles are also continually being consumed by different organisms throughout the mesopelagic and either disaggregated or re-combined into different fecal pellets (Poulsen and Kiørboe, 2005; Wilson et al., 2008; Robinson et al., 2010). Incorporating such transformations into conceptual and numerical models of particle flux through the euphotic zone is needed to explain and predict changes in the ocean's remineralization length scale and ability to sequester carbon dioxide (Iversen and Poulsen, 2007; Poulsen and Iversen, 2008; Kwon et al., 2009; Lomas et al., 2010; Burd et al., 2016).

CHAPTER 5

CONCLUSION

Our paired measurements of particle size and volume using UVP optical measurements compared with carbon flux determined with sediment traps and ^{238}U - ^{234}Th disequilibrium clearly show that previously published algorithms for estimating particle flux from UVP data do not perform well in the CCE. This adds to a growing body of literature suggesting that particle size-flux relationships are highly variable in the ocean and that such variability must be taken into account when applying optically derived estimates of flux. A CCE-optimized parameterization leads to a relatively low exponent ($B = 1.05$) for the size-flux relationship. This low exponent likely results from the important role of 100-400 μm fecal pellets in vertical carbon flux in the CCE, a carbon source that is poorly resolved in optical profiles. Further improvements in optical estimates of carbon flux will require imaging systems with a higher resolution at small particle size, the ability to discriminate between different classes of particles (e.g., fecal pellets and marine snow), and accurate representation of rapidly sinking fecal pellets and other sources of organic matter. Until such advancements are made it is advisable that future UVP derived estimates be paired with other direct methods of flux estimation as has previously been suggested.

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BIOGRAPHICAL SKETCH

CHRISTIAN KADE FENDER

EDUCATION

Florida State University; Tallahassee, FL

PhD in Biological Oceanography, currently pursuing (August 2018 – present)

Florida State University; Tallahassee, FL

Masters of Science in Chemical Oceanography, (November 2022)

University of South Florida; Tampa, FL

Bachelors of Science in Marine Biology, (May 2018)

Honors College and Provost's Scholar

RESEARCH EXPERIENCE

Undergraduate:

- Gemmell lab: Thesis on the impact of environmental stressors on pulse rate of the upside-down jellyfish *Cassiopea sp.*, University of South Florida– Tampa, Florida (2017-2018)

Graduate:

- Carbon Export Modelling with Optics (June 2018 - October 2019)
 - Explored the use of cutting edge underwater cameras to estimate particle export in the California Current and compare to traditional sediment trap and ²³⁴Th methods.

- Gelatinous Zooplankton Ecology and Biogeochemistry (October 2018 – present)
 - Investigating the trophic role of salps in New Zealand’s Chatham rise with an emphasis on the bioenergetic implications of their prey size, predator:prey ratios, and selection for specific taxa
 - Future work will explore the efficiency with which fecal pellets, sinking corpses, and vertical migration export carbon to depth in a 3D physically coupled model.

- Bluefin Larvae in Oligotrophic Ocean Foodwebs, Investigations of Nutrients to Zooplankton (Jan. 2021 – present)
 - Ongoing work to study the nutrient landscape and trophic structure of the Argo Basin to determine its impact on the Indian Ocean spawning grounds of Eastern Bluefin Tuna.

- ICER NNA – Navigating the New Arctic (spring 2023)
 - Will be assisting in the tuning of a specially designed version of the NEMURO ecosystem model which will be coupled to a 3D HYCOM physical model.
 - Outputs of the lower trophic order organisms and how they respond to climate change will be used to estimate the resilience of the Maine lobster fishery and inform partners in the management of both coastal New England and Atlantic Canada.

PEER REVIEWD PUBLICATIONS

Fender, C. K., M. Decima, A. Gutierrez-Rodriguez, K.E. Selph, N. Yingling, M. R. Stukel (in review). Prey Size Spectra and Predator:Prey ratios of 7 Species of New Zealand Salps. *Marine Biology*.

Fender, C. K., Kelly, T. B., Guidi, L., Ohman, M. D., Smith, M. C., and Stukel, M. R. (2019). Investigating Particle Size-Flux Relationships and the Biological Pump Across a Range of Plankton Ecosystem States From Coastal to Oligotrophic. *Front. Mar. Sci.* 6, 603. doi:10.3389/fmars.2019.00603.

CONFERENCE PRESENTATIONS AND ABSTRACTS

Fender, C. K., M. Decima, A. Gutierrez-Rodriguez, K.E. Selph, N. Yingling, M. R. Stukel (2021). VISUALIZING THE VACUUM: EVIDENCE FOR NICHE PARTITIONING IN NEW ZEALAND SALPS. In *2021 Aquatic Sciences Meeting*. ASLO.

Décima, M., Stukel, M. R., Selph, K. E., Gutierrez-Rodriguez, A., Safi, K., Nodder, S. D., Luskow, F., **Fender, C.**, Meyers, M. (2020). Salp Grazing Effects on the Lower Food Web and Carbon Export, in Subtropical and Subantarctic Waters East of New Zealand. In *Ocean Sciences Meeting 2020*. AGU.

Fender, C. K., T. B. Kelly, L. Guidi, M. D. Ohman, M. C. Smith, M. R. Stukel (2020). Investigating Particle Size-Flux Relationships and the Biological Pump Across a Range of Plankton Ecosystem States From Coastal to Oligotrophic. In *2020 Southeastern Biogeochemistry Symposium*.

PARTICIPATION IN EDUCATION

- Guest Lecture for Dr. Oliva Mason. Lecture titled: *OCB-5636 Intro to Graphing and Figure Editing in R*. November, 2015.
- Volunteer Outreach Scientist at COAPS Annual Open House. Lesson – *Plankton do the strangest things*. February, 2018.

- Volunteer Outreach Scientist at FSU Coastal Marine Lab Annual Open House. Lesson – *Gelatinous Zooplankton*. April, 2022.
- Teaching Assistant, Introduction to Environmental Science Lab (EVR1001L), Florida State University (2018-2021)
- Teaching Assistant, Introduction to Environmental Science (EVR1001), Florida State University (Fall Semester, 2021)
- Teaching Assistant, Biological Science 2 Lab (BIO2011L), Florida State University (Spring Semester, 2022)
- Teaching Assistant, Biological Science 1 Lab (BIO2010L), Florida State University (Fall Semester, 2022)

CRUISE EXPERIENCE (total ship-time: 150 days)

- BLOOFINZ 2022 – Determined carbon export via ^{234}Th : ^{238}U disequilibrium and drifting surface tethered sediment traps, including traps to determine transcriptional activity of sinking particles. Also deployed various nets to determine size fractionated zooplankton abundance and measured chlorophyll concentrations and HPLC pigments from in situ drifting arrays.
- CCE-P2107 - Carbon export using Th and sediment traps to determine new production in a coastal filament as part of the California Current Ecosystem Long Term Ecosystem Research project.
- CCE-P1908 - Measured carbon export, new production, and zooplankton biomass in a coastal jet in the CCE.
- SalpPOOP 2018- An NSF funded project in collaboration with NIWA to study the biogeochemical role of Salp fecal pellets and the ecological impact of salp blooms. Conducted carbon particle measurements on the cruise including sediment traps and thorium measurements. Deployed a variety of nets to sample zooplankton abundance, determine salp grazing, and image salp gut contents using Scanning Electron Microscopy. Also responsible for the preparation and analysis of ^{13}C , ^{15}N , and

compound specific isotopic analysis of amino acids of zooplankton and salp biomass samples to determine trophic order.